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SIMULTANEOUS SYNTHESIS OF BOTH DIASTEREOMERS OF STEREOSELECTIVELY β -DEUTERATED PHENYLALANINES:(2S,3R) - AND (2R,3R)-PhCHDCH(NH₂)COOH

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Precise ¹H NMR analysis of side chain conformation of amino acids having a β -methylene group requires discrimination of the two prochiral protons on the β -carbon.^{1,2} It can be achieved by stereoselective deuteration. Stereoselective β -deuteration of several amino acids by chemical^{3,4} and enzymatic⁵ methods are published. Kirby and Michael³ prepared (2S,3R)-phenylalanine-3-d₁ by catalytic hydrogenation of <u>trans</u>- α -acylamidocinnamic acid-3-d₁ and subsequent optical resolution. The method affords only one of the two possible diastereomers. Single diastereomer of known relative configuration suffices to discriminate the two β -protons when their signals are well separated in the NMR spectrum.² However, use of both diastereomers is desired for an unambiguous assignment when the two β -proton signals are in close proximity. Thus, we have designed the synthesis of both of the diastereomers as follows.

Reduction of benzaldehyde- α -d₁ (1) prepared from phenylglyoxylic acid by actively fermenting yeast afforded (S)-benzyl alcohol- α -d₁ (2),⁶ whose d-content



was 93% (NMR) and the optical purity was confirmed by NMR using a chiral shift reagent, tris[3-(heptafluoropropylhydroxymethylene)-<u>d</u>-camphorato]praseodymium, according to the reported procedure.⁷ The methylene proton signal appears as a broad singlet corresponding to the pro-R proton signal (at a higher field) of the two methylene proton signals of benzyl alcohol (0.13 M in CCl₄) in the presence of 0.3 molar ratio of the chiral shift reagent. Tosylate (3) reacted with ethyl sodioacetoacetate to give α -benzylacetoacetate (4) with inversion of the configuration as discussed later.⁸ Treatment of <u>4</u> with hydrazoic acid under the standard condition of Schmidt reaction gave N-acetyl-DL-phenylalanine-3-d₁ (5). Hydrolysis of <u>5</u> by α -chymotrypsin⁹ afforded the desired products, (2S,3R)-N-acetyl-L-phenylalanine-3-d₁ (6) and (2R,3R)-N-acetyl-D-phenylalanine-3-d₁ ethyl ester (7) in 44% and 45% yield, respectively. This is the first synthesis of a (2R,3R)-phenylalanine derivative.

The optical purities of <u>6</u> and <u>7</u> were determined by molecular rotation ([ϕ]) at the trough (225 nm). Their [ϕ]₂₂₅-values were +13200° for <u>6</u> and -11600 for <u>7</u> (c=1.25% in MeOH), which indicated the 91% and 94% purity, respectively, by comparison with the standard samples. The diastereomeric purity of <u>7</u> was determined by ¹H NMR in CDCl₃ with the aid of a shift reagent, Eu(dpm)₃, which separates the two β -methylene signals; the pro-R proton signals of the D-Phe derivative appear at a higher field. An intensity ratio of 83:17 observed corresponds to the epimeric purity of 92% at C-3 (calculated from the d-content and optical purity at C-2). The epimeric purity should be equal to that for <u>6</u> in consideration from the synthetic route. This result implies that almost complete inversion of the configuration was caused in the carbanionic reaction and the tosylation step.

The overall yield was about 6 for both <u>6</u> and <u>7</u>, which is almost equal to that with the Kirby's procedure.³ Our method takes two steps longer, but afford both diastereomers. The loss of deuterium was not observed through all the steps

Studies on a rotameric equilibrium of the side chain in phenylalanine derivatives are in progress using derivatives of <u>6</u> and <u>7</u>. These compounds will also be useful for stereochemical studies on biogenesis including phenylalanine as a substrate or a product.

REFERENCES

- 1) G.C.K. Roberts and O. Jardetzky, Advan. Protein Chem. 24, 477 (1970)
- 2) M. Kainosho and K. Ajisaka, <u>J. Amer. Chem. Soc</u>. <u>97</u>, 5630 (1975)
- 3) G.W. Kirby and J. Michael, <u>J. C. S. Perkin I</u> 115 (1973)
- 4) P. Stetter, Jr., J. Biol. Chem. 144, 501 (1942)
- 5) H.J. Bright, R.E. Lundin and L.L. Ingraham, Biochemistry 9, 1224 (1964)
- 6) V.E. Althouse, D.M. Feigl, W.A. Sanderson and H.S. Mosher, <u>J. Amer. Chem. Soc</u> <u>88</u>, 3595 (1966)
- 7) R.R. Fraser, M.A. Petit and M. Miskow, <u>Ibid</u>. <u>94</u>, 3253 (1972)
- A. Streitwieser, Jr., J.R. Wolfe, Jr. and W.D. Schaeffer, <u>Tetrahedron</u> <u>6</u>, 338 (1959).
- 9) M.S. Matta, J.A. Kelley, A.J. Tietz and M.F. Rohde, J.Org. Chem. 39, 2291 (1974

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